It is necessary to see that the strainer does not become choked. This may be prevented by shaking. The strainer should, of course, be kept in water between uses. When it finally becomes clogged with sand, a new one must be put on.

By using wood with different sized conducting vessels, a sorting of the diatoms may be affected. By using pine, spruce, white wood of the red cedar, a graded series of strainers can be had, the last being much finer than the first.

STAINING PROTOZOA

Darling (Science: Jan. 10, 1913) calls attention to the dearth of knowledge of the acidophilic substances in the nuclei of protozoa, owing to the predominant use of basophilic staining substances, and to the "lack of a satisfactory technic for demonstrating acidophilic substance in wet fixed films."

The author suggests careful differentiation of such polychrome stains as Romanowsky or Hastings-Giemsa, by ammoniated ethyl alcohol. Under such conditions, studying *Entamœbæ,* he found a definite arrangement of an acidophilic substance (oxychromatin) within the nucleus, showing a structure quite different from that shown with the usual basichromatin stains. He believes that careful and critical study will reveal that this oxychromatin may have important functional relations to the changes that are so well known in the true chromatin in nuclear activity.

DOUBLE-STAIN METHOD FOR THE POLAR BODIES OF DIPHTHERIA BACILLI

Dr. Marie Raskin (Apoth. Ztg. XXVII, p. 10; Abstr. United St. Naval Med. Bull. Vol. 6, No. 4, p. 611) proposes a technic for these bodies, whose distinctive value lies in the fact that only two operations are necessary, i. e., the application of a stain with both colors present, then water washing.

Formula for stain:—

Glacial Acetic Acid.....	5 cc.
Dist. Water	95 "
Alc. (95%)	100 "
Old Sat. sol Methylene blue.....	4 "
Ziehl's phenol fuchsine Sol.....	4 "

Drop mixture in a thin layer over the specimens on the cover glass; heat through the flame. The alcohol ignites and is permitted to burn off, after which the specimen is washed in water and dried. The entire process takes 20-25 seconds, and the stain remains serviceable for any length of time. Polar bodies appear deep blue and the bacilli bright red. Even in smears with a preponderance of other bacteria, individual diphtheria bacilli may be readily and unmistakably identified.

A NEW TECHNIC IN STAINING DIPHTHERIA SPECIMENS WITH TOLUIDIN BLUE

Dr. Constant Ponder (Lancet, July 6, 1912; Abstr. U. S. Naval Med. Bull. Oct. 1912, p. 612) recommends the following treatment for diphtheria bacilli:—

The stain:

Toluidin blue (Grübler)	0.02 gram.
Glacial acetic acid	1 cc.
Abs. Alc.	2 "
Distilled Water to make	100 "

The film made on cover glass is fixed as usual. Spread stain on film. The cover glass is then turned over and mounted as a hang-drop preparation. Typical diphtheria bacilli are said to stain blue, with red granules. The author gives this as a new method, and says it is preferable to either Methylene blue or Neisser's stain.

NOTES FROM MEETING OF THE ILLINOIS MICROSCOPICAL SOCIETY, *Chicago, Oct. 10, 1912*

Mr. N. S. Amstutz showed a useful contrivance for keeping pond life in place. It consisted of a piece of brass about 7/8 in. square and 5/32 in. thick. A series of seven holes were drilled thru it so as to imprison that many varieties of pond life at one time. The plate was placed in a flat bottomed watch glass and each specimen transferred with a pipette to its proper "cell." These could be then studied at will very nicely with a 2/3 objective and various combinations of oculars. The specimens were confined laterally so they were unable to move out of the field of view though having abundant room for vertical movement. With the coarse ad-